



Identification of bisphenol A diglycidyl ether derivatives after *in vitro* digestion process by LC-MS/MS

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1. INTRODUCTION

Epoxy-based resins, obtained by polymerization between bisphenol A (BPA) and epichlorohydrin, are well-known compounds used for **coating** applications in food metal cans [1]. This chemical group has arisen as a public concern because of the **endocrine disruptor** character of their monomer, BPA, along with the possible health effects that their major derivatives may pose on consumers [2].

For a correct estimation of the exposure, the **bioaccessibility** must be considered, this means, the fraction of the contaminant available to be absorbed. With that purpose, the INFOGEST protocol [3] was applied.

Objectives

- → Perform an accelerated **stability test** to simulate the storage of tuna cans.
- → Study the gastrointestinal fate of bisphenol A diglycidyl ether (BADGE), one of the main contaminants related to the cited resins.
- → Identify the transformation products of BADGE, with water and chloride by liquid chromatography coupled to tandem mass spectrometry (LC-MS/MS).



2. MATERIAL AND METHODS

2.1. Stability test	2.2. In vitro digestion		2.3. Identification with LC-MS/MS					
	INFOGEST protocol Gastrointestinal digestion through three phases testing the influence of gastric pH	Table 1. Experimental conditions of HPLC-FLD and LC-MS/MS						
Olive oil: commonly used covering liquid in cans		Equipment	LC-MS TSQ Quantum Access MAX and Agilent Technologies	P	Phase mobile gradient			
Canned tuna: greatly	gasme pri		1100 series HPLC	Time (min)	ACN (%)	H ₂ O (%)		
consumed product with high protein content	SSF: simulated salivary fluid	Column	Phenosphere 80Å ODS (150 mm x 4.6 mm, 3µm)	Ο	40	60		
	SGF: simulated gastric fluid SIF: simulated intesting! fluid	Mobile phase	ACN:H2O	3	40	60		
		Flow rate	1mL/min	19	75	25		
			,	24	100	0		
Food matrixes		Injection volume	10µL	26	100	0		
spiked with BADGE	1. Oral phase	Fluorescent detection	Excitation: 225 nm Emission: 305 nm					
	5 g sample, amylase SSF, 2 min pH 7	MS data acquisition	Selected reaction monitoring (SRM) and Full Scan					
60° for 10 days, according to (EU) No	2. Gastric phase Pepsin, HCl SGE 2 hr pH 123 and 4	Source	Positive and negative atmospheric pressure chemical ionization (APCI)					
	7 Intecting phase	Spray voltage	2500 V	Theme Access				
	Dile server l'e Ne Old	Vaporizer T ^o	340°C		. 7			
intended to come into contact with food [4].	Sile, pancreatin, NaOH SIF, 2 hr pH 7	Capillary T°	350°C	Figure 1. LC-MS/MS 1 Scientific	Thermo Fig	ure 2. Triple quadrupole and data acquisition diagram		

3. RESULTS AND DISCUSSION

Compound	CAS	APCI mode	Parent ion	Product ions	Collision energy (V)
BADGE·2H ₂ O	5581-32-8	-	421.4	226.8	30
				300.6	17
BPA	80-05-7	-	227.0	134.5	29
				213.1	22
BADGE·HCl·H ₂ O	227947-06-0	-	439.4	82.5	14
				393.5	15
BADGE·H ₂ O	76002-91-0	+	400.0	107.1	41
				135.1	25
BADGE-2HCI	4809-35-2	-	457.0	82.4	15
				393.2	21
BADGE·HCI	13836-48-1	+	418.0	106.9	40
				135.1	28
BADGE	1675-54-3	+	382.3	135.1	25
				191.1	19
CycloDiBADGE	20583-87-3	+	569.0	135.1	27
				107.1	43

Table 2. Parameters of identification of the standard mix using LC-MS/MS





Figure 4. LC-MS/MS chromatogram using negative APCI ionization mode of digested olive oil spiked with BADGE



Figure 5. LC-MS/MS chromatogram using positive APCI ionization of digested olive oil spiked with BADGE

BADGE-2H₂O was identified after analyzing the bioaccessible fraction of the sample of olive oil spiked with BADGE using negative APCI as ionization mode.

BADGE and BADGE·HCl were found in the bioaccessible fraction of the sample of olive oil spiked with BADGE using positive APCI as ionization mode.

4. CONCLUSIONS

Liquid chromatography coupled to tandem mass spectrometry (LC-MS/MS) allowed the identification of BADGE and its derivatives in the samples obtained after the *in vitro* digestion process.
The preliminary results showed that the studied chemical, BADGE, reacts with water and chloride during gastrointestinal digestion to form BADGE:2H₂O and BADGE:HCl, respectively.
These compounds have been identified in the bioaccessible fraction of the spiked olive oil, one of the most common covering liquids used in canned food.
BADGE derivatives present in the standard mix were not found in tuna samples. The possible biotransformation of BADGE in presence of this food matrix will require more investigation.
Further research should be undertaken to study the effect of different food matrices on the transformation of BADGE, together with the tentative identification of other substances yet unknown.

5. REFERENCES

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