

## IDENTIFICATION OF POTENTIAL MIGRANTS PRESENT IN POLYMERIC COATINGS IN FOOD CANS

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## INTRODUCTION

Polymeric coatings are commonly used in metal food cans to protect food from corrosion. However, there is a risk that coatings used in cans release complex chemical mixture into foodstuff including monomers, oligomers, additives, impurities, reaction products, etc. It is important to develop analytical tools for the identification of these potential migrants in the food packaging with the ultimate objective of ensure the consumer safety. A GC-MS screening method was performed to identify potential migrants in polymeric coatings and a LC-MS/MS method was developed for the determination of bisphenols and BADGEs. A total of twelve food cans that cover several types of food including fish, seafood, vegetables and fruit were selected as study samples.

## EXPERIMENTAL

Identification of the type of coating (ATR-FTIR)

Identify the type of coating, infrared spectra were acquired using an ATR (attenuated total reflectance)-FTIR spectrometer equipped with a diamond optical crystal. The spectra identification was performed by comparing recorded spectra with several commercial spectral libraries (IR Spectral Libraries of Polymers & Related Compounds from Bio-Rad Laboratories).



Can sample



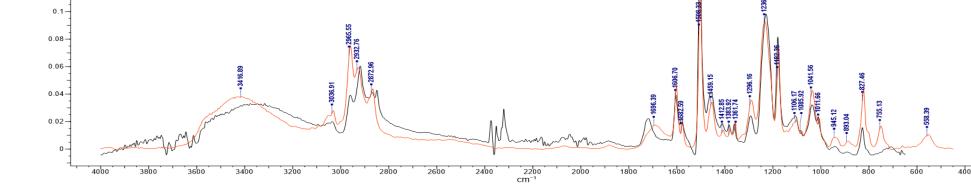


Figure 1. IR spectrum of the internal side of one sample (dark) compared to the IR of the Spectral Library (red)

### Identification of potential migrants (GC-MS)

# A non-targeted screening by gas chromatography coupled to mass spectrometry (GC-MS) was performed for migrant identification.

					Column	ZB-5MS (30 m × 0.25 mm × 0.25 μm)
					Carrier gas	Helium 1mL/min
					Injection mode	Splitless
					Injection volume	1μL
					T <sup>a</sup> program	40ºC/2min, 9ºC/min to 300ºC, 300ºC/3min
	and the construction of th			Thermo man and an Thermon There are	Data acquisition	Full scan (range m/z 35-500)
Can sample	A known surface of the	An aliquot of the extracts	An aliquot was		Mass detector	Electron impact
Gan Gampio	packaging was put in contact with a volume of acetonitrile for 24 h at 70∘C	(10 mL) was evaporated down to 1 mL by a stream of nitrogen	filtered and analysed by GC-MS		Transfer line T <sup>a</sup>	300ºC
					Detector T <sup>a</sup>	300ºC
					Mass spectrum library	Wiley 8th & NIST/EPA/NIH 11

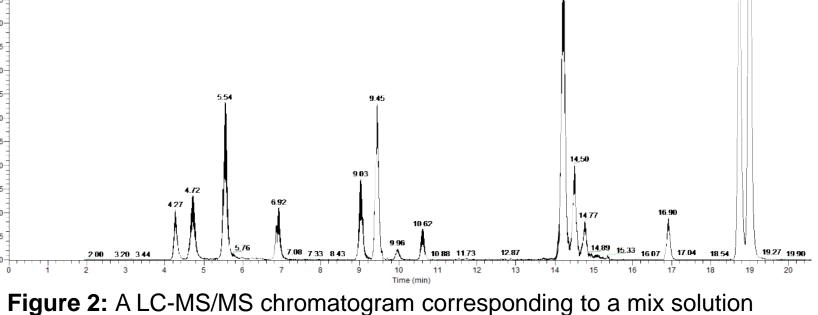
# Multi-target analysis (LC-MS)

A targeted analysis was carried out for the simultaneous determination of thirteen compounds including bisphenols and BADGEs by liquid chromatography coupled to tandem mass spectrometry (LC-MS/MS).

There Area	

		Phenosphere 80A ODS	Compound	Retention Time (min)	APCI mode	Parent ion	Product ion	Collision energy (V)	RT: 0.00 - 20.64 SM: 7B	
	Column	$(150 \text{ mm} \times 3.2 \text{ mm}, 3 \mu\text{m})$	BPF	4.27	-	198.9	93.0 105.0	24 23	100 95-	
	Mobile phase	MeOH: ACN (50:50, $v/v$ ) and water	BADGE.2H <sub>2</sub> O	4.72	-	374.8	226.8 300.6	28 16	90	
	Flow rate	0.5 mL/min	BPE	5.54	-	212.9	196.8 197.8	33 20	85	
	Injection volume	10μL	ВРА	6.92	-	226.9	133.0 211.8	28 20	75	
		55% water and 45% MeOH:ACN for 2min,	BPB	9.03	-	240.9	210.7 211.8	31 20	65-	
10 TSO Guartian Access MAX		MeOH:ACN was increasing until 75% for	BADGE.H <sub>2</sub> O	9.45	+	399.9	106.9 134.8	45 26	60- 9 55- 9 55-	
	Gradiente elution	14min, and another gradient to 100%	BADGE.H <sub>2</sub> O.HCl	9.96	-	283.0	211.0 226.0	30 21	In 50	
		MeOH:ACN for 7 min	BPC	10.62	-	254.9	146.9 239.8	33 21	<u>د</u> 40	
	Data acquisition	Selected reaction monitoring (SRM)	BADGE	14.23	+	381.9	134.9 190.8	31 25	30-	5.54
		Positive and negative atmospheric	BADGE.HCI	14.50	+	417.9	106.9 134.9	43 28	25-	
	Source	pressure chemical ionisation (APCI)	BADGE.2HCl	14.77	+	382.2	191.1 135.2	16 26		
	Vaporizer Tª	400°C	BPG	16.90	-	311.0	174.9 294.9	33 37	10- 5-	
	· · · · · ·	350°C onditions of LC-MS/MS method	CYDBADGE	18.74 <i>,</i> 18.99	+	569.0	134.8 106.9	29 39	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	5 6
	Table 2: Experimental c	onaltions of LC-IVIS/IVIS method							Figure 2: A   C-MS	2/1/2





#### Table 3: MS/MS conditions and retention times

## **RESULTS AND DISCUSSION**

The FTIR results confirmed that most of the samples examined in this study were coated with epoxy-phenolic resins.

The screening reveals the presence of a great variety of compounds including plasticizers (acetyl tributyl citrate, bis(2-ethylhexyl) adipate, phthalates such as diisobutyl phthalate, bis(2-ethylhexyl)phthalate, etc.), photoinitiators (benzophenone and 4-phenylbenzophenone), antioxidants (butylated hydroxytoluene), lubricants (butyl palmitate, glycerol tricaprylate), etc. Sixteen of the total of forty-five compounds were positively confirmed by injection of the respective standard. Several degradation products formed from antioxidants used as additives were identified in the can extracts, for example, 1,3-di-tert-butylbenzene and 2,4-di-tert-butylphenol.

The developed method by LC-MS/MS turned out to be an excellent analytical tool for the confirmation of the presence of bisphenol related compounds in the can extracts being cyclo-di-BADGE the predominant compound detected.

Compound	Samples											
Compound	ΑΑ	AH	AL	AN	AR	ES	MA	ME	MZ	SR	TO1	TO2
BPF												
BADGE.2H <sub>2</sub> O	Х	Х	Х	Х		Х	Х	Х	Х	Х	Х	Х
BPE												
BPA				Х		Х		Х				Х
BPB												
BADGE.H <sub>2</sub> O	Х	Х	Х	Х		Х	Х	Х	Х	Х	Х	Х
BADGE.H <sub>2</sub> O.HCl			Х	Х				Х				
BPC												
BADGE		Х	Х	Х		Х	Х	Х	Х	Х	Х	Х
BADGE.HCI		Х	Х	Х								Х
BADGE.2HCI												
BPG												
CYDBADGE	Х	Х	Х	Х	Х	Х	Х	Х	Х	Х	Х	Х

**Table 4:** Bisphenols and BADGEs identified in the extracts of the analysed cans by LC-MS/MS



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