

A GC-MS METHOD FOR IDENTIFICATION OF POTENTIAL CONTAMINANTS IN FOOD PLASTIC PACKAGING AND A GC-MS METHOD FOR DETERMINATION OF CONTAMINATS IN FOODSTUFFS

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INTRODUCTION

The safety of food packaging materials is a cause of concern since they can transfer chemical compounds into the food which may affect not only the safety but also the quality of the packaged food. In this work, the potential migration of chemicals from plastic food packaging into the food was investigated. A total of fourteen samples of dairy products were selected for this study and a GC-MS method was developed to quantify selected compounds in the foodstuffs. The method was validated in terms of linearity, repeatability and recoveries.

EXPERIMENTAL

Fourteen samples of dairy products comprising milk, yogurt, flan and cheese, were purchased in a local supermarket of Santiago de Compostela. Labeling detailed information about each food was recorded, including fat/saturated fat (ranged from 1.6% to 35%). To verify the type of material, infrared spectra were acquired using an ATR (attenuated total reflectance)- FTIR spectrometer equipped with an optical crystal of diamond. The spectra identification was possible by comparing recorded spectra with several spectral libraries related with polymers and additives.

	Coding	Type of comple	Fat	Part of the comple	Type of material			
	Coung	Type of Sample		Part of the sample	Internal side	External side		
	LE_01	Whole milk	3.6 %	Packaging	Polyethylene	Polyethylene		
	LE_02	Whole milk	3.6 %	Packaging	Polyethylene	Polyethylene		
	LE_03	Whole milk	3.6 %	Packaging	Polyethylene	Polyethylene Polyethylene Polyethylene		
	LS_01	Semi-skimmed milk	1.6 %	Packaging	Polyethylene			
	LS_02	Semi-skimmed milk	1.9 %	Packaging	Polyethylene			
	YN_01	Natural yogurt	2.9 %	Lid	Polyethylene terephthalate	Nitrocellulose		
				Packaging	Polystyrene	Polystyrene		
	VS 01	Strawberry yogurt	1.9 %	Lid	Polyethylene terephthalate	Nitrocellulose		
				Packaging	Polystyrene	Polystyrene		
	FN_01	Egg flan with caramel	1.8 %	Lid	Styrene-butadiene-isoprene rubber	Nitrocellulose		
				Packaging	Polypropylene	Polypropylene		
	FN_02	Egg flan	3.2 %	Lid	Based in Methacylate	Based in Acrylate		
				Packaging	Copolymer vinyl chloride/vinyl acetate	Epoxy resin		
		Custard	3 %	Lid	Polyethylene terephthalate	Nitrocellulose		
				Packaging	Polystyrene	Polystyrene		
		Semi-cured cheese	35 %	Lid	Polyethylene	Polyethylene terephthalate		
	QS_01			Packaging	Polyethylene	Polyethylene terephthalat		
				Intermediate sheet	Polystyrene	Polystyrene		
	01 01	Molton chaoso	13.5 %	External packaging	Polyethylene	Polyethylene terephthalate		
		Mollen cheese		Internal packaging	Polyethylene	Polypropylene		
	QF_01	Mozzarella	18 %	Packaging	Polyethylene	Nylon (Polyamide)		
	OF 02	Pastourized chaose	14 %	Lid	Polyethylene terephthalate	Based in Polyurethane		
		i asteunzeu cheese		Packaging	Polystyrene	Polystyrene		



Table 1: Information about the samples of the study.

SCREENING OF THE POTENTIAL MIGRANTS IN FOOD PACKAGING MATERIALS

In a first step, a gas chromatography with mass spectrometry (GC-MS) method was developed to identify potential migrants in an extract of the packaging.



SELECTED MIGRANTS OF JANTIFICATION IN THE FOODSTUFFS

In the <u>second step</u> of the work, the packaged foods were pooled into three groups according to the population age (12-35 months, 3-9 years and 10-17 years) based on the Spanish consumption data (Enalia) and analyzed by GC-MS to quantify



	500-C, 500-C/ 5mm				
Data acquisition	Full scan (range m/z 35-500)				
Mass detector	Electron impact				
Transfer line T ^a	300ºC				
Detector T ^a	300ºC				
	Wiley 8th & NIST/EPA/NIH 11 Mass				
Spectrum library	spectral library (version 2.0)				

Table 2: Experimental conditions of GC-MS method.

RESULTS AND DISCUSSION

Vortex shake Filter an aliquot and analysis by GC-MS

Evaporation to dryness using a stream of nitrogen

The experimental conditions of GC-MS method are the same as in the food packaging, except some differences: the injection mode split (1:5), the initial T^a in the oven is 60°C and the data acquisition is in mode SIM.

More than 90 compounds were identified in the food packaging and selected compounds were confirmed using standards. Among them, antioxidants as butylated hydroxytoluene (BHT); plasticizers as diethyl phthalate (DEP), diisobutyl phthalate (DIBP), acetyltributyl citrate (ATBC), bis(2-ethylhexyl)phthalate (DEHP), bis(2-ethylhexyl) terephthalate (DEHT), diethylhexyl adipate (DEHA); photoinitiators as benzophenone (BP); thermal decomposition product of polystyrene as 1,3-diphenylpropane (1,3-DPP); UV filters as octocrylene; and slip agents as hexadecanamide and erucamide.

Results of this study demonstrate the migration of phthalates from food packaging materials to foodstuffs, in concentrations that range from 0.02 μ g/g of DEP to 0.16 µg/g of DBP (pool 10-17 years). All compounds present good linearity in the studied quantification range ($R^2 \ge 0.9900$). Appropriate recovery range at the three spiking levels (from 78.4 % to 124.4 %) and a good sensitivity was obtained with this method.

Commonwed	Sample	LOD µg/g) (µg/g)	LOQ (µg/g)	Recovery (%)		Repeatability (RSD %)			
Compound	Concentration (µg/g)			0.25 μg/g	0.5 µg/g	1 μg/g	0.25 μg/g	0.5 µg/g	1 µg/g
DEP	0.02-0.03	0.01	0.025	112.0	113.9	102.2	16.1	8.36	2.50
BP	<lod< th=""><th>0.05</th><th>0.1</th><th>107.1</th><th>103.6</th><th>102.1</th><th>14.9</th><th>7.67</th><th>2.03</th></lod<>	0.05	0.1	107.1	103.6	102.1	14.9	7.67	2.03
1,3-DPP	<lod< th=""><th>0.025</th><th>0.05</th><th>88.8</th><th>80.0</th><th>78.4</th><th>5.03</th><th>1.73</th><th>7.01</th></lod<>	0.025	0.05	88.8	80.0	78.4	5.03	1.73	7.01
DIBP	0.05-0.10	0.005	0.005	112.1	123.1	124.4	5.40	2.80	1.61
DBP	0.09-0.16	0.005	0.005	119.8	123.9	123.0	8.52	0.98	4.26
DEHP	0.07-0.09	0.005	0.005	122.7	123.9	115.8	1.62	1.04	7.51
DEHT	<lod< th=""><th>0.025</th><th>0.05</th><th>96.2</th><th>89.6</th><th>98.9</th><th>16.0</th><th>15.9</th><th>18.2</th></lod<>	0.025	0.05	96.2	89.6	98.9	16.0	15.9	18.2

Table 3: Sample concentrations and method validation parameters.



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